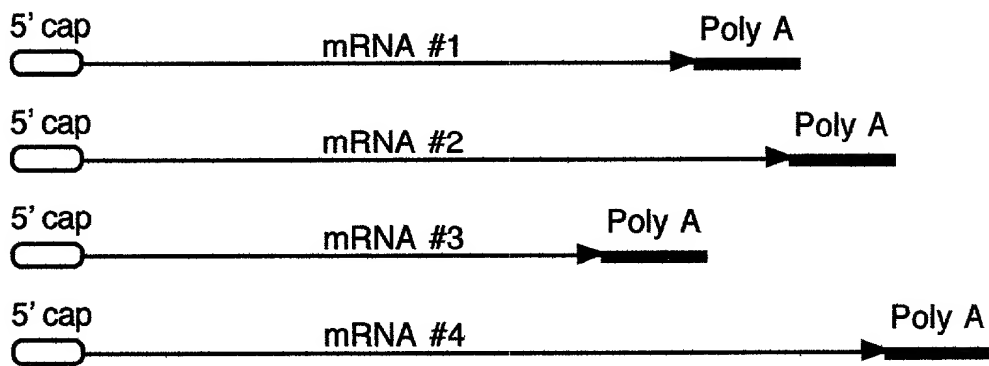


Library of mRNA analytes



Library of mRNA analytes bound to an array

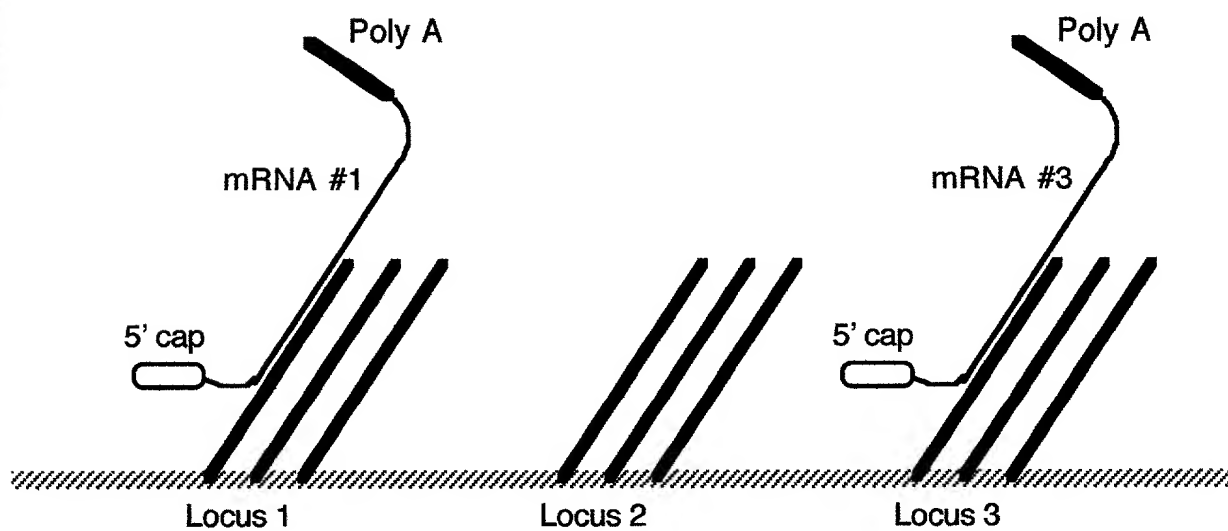


Figure 1

A) RNA substrate



B) Fragmentation of RNA substrate



C) addition of tails (UDTs) to RNA fragments



D) Detection of tails (UDTs) on RNA fragments by binding a reagent containing signal groups (S)

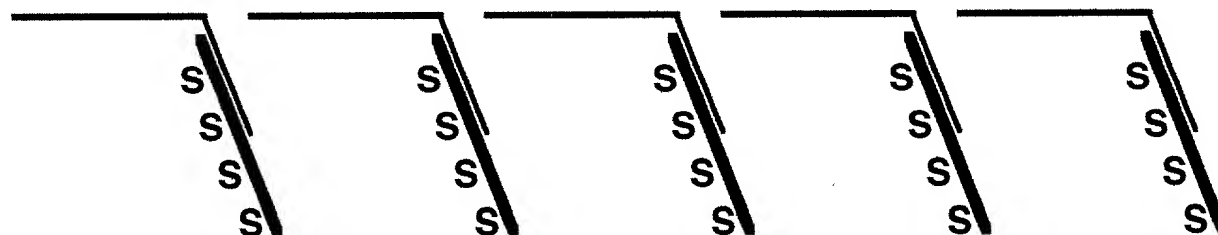
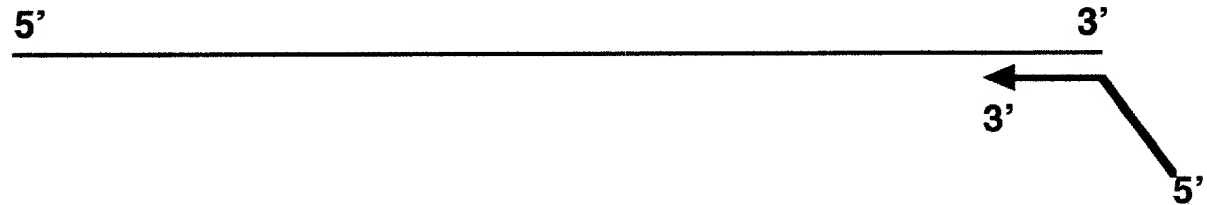


FIGURE 2

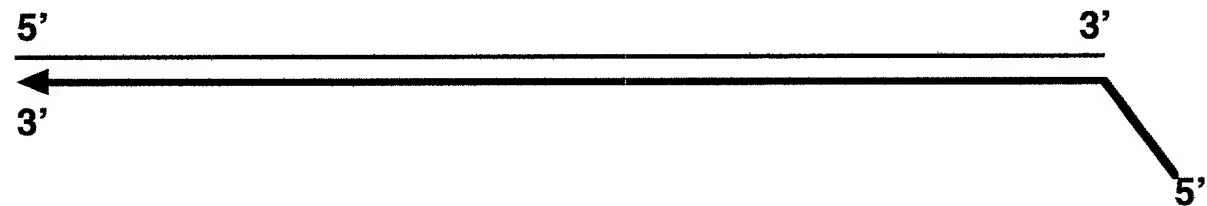
(A) RNA Substrate



(B) Binding of Primer to RNA Substrate



(C) Extension of Primer using RNA as template



(D) Template Independent Extension of Primer

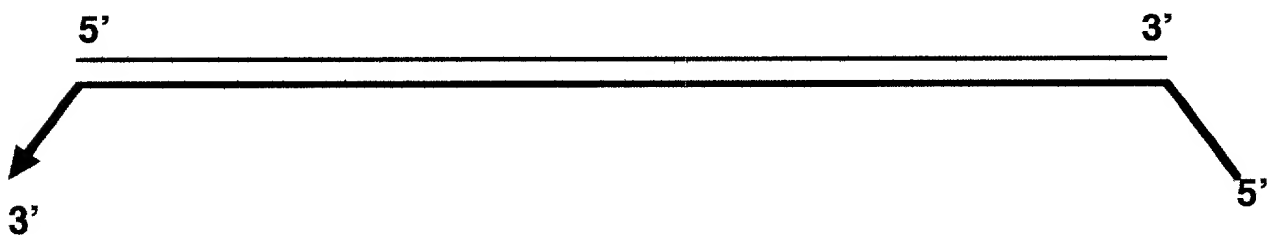
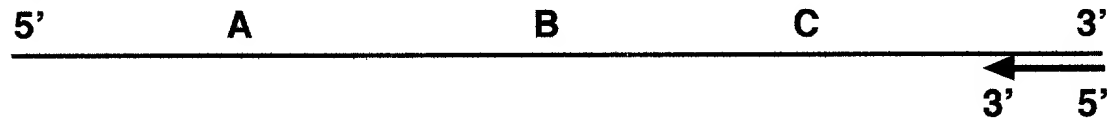


FIGURE 3

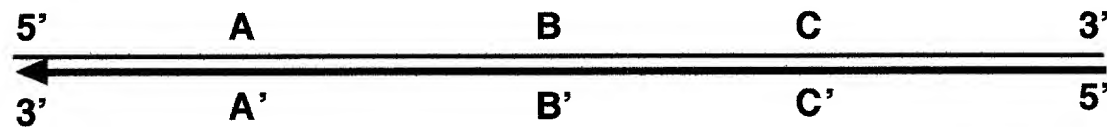
(A) RNA Substrate



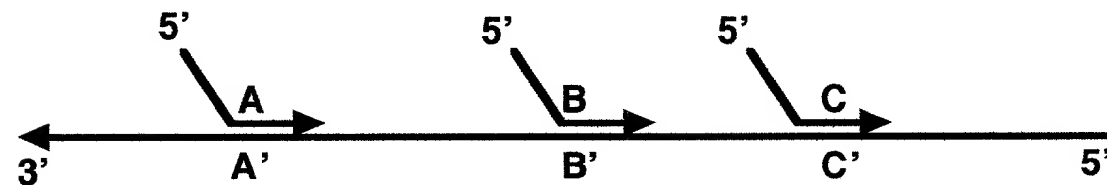
(B) Binding of Primer to RNA Substrate



(C) Extension of Primer using RNA as template



(D) Binding of random primers to 1st cDNA strand



(E) Extension and strand displacement of random primers

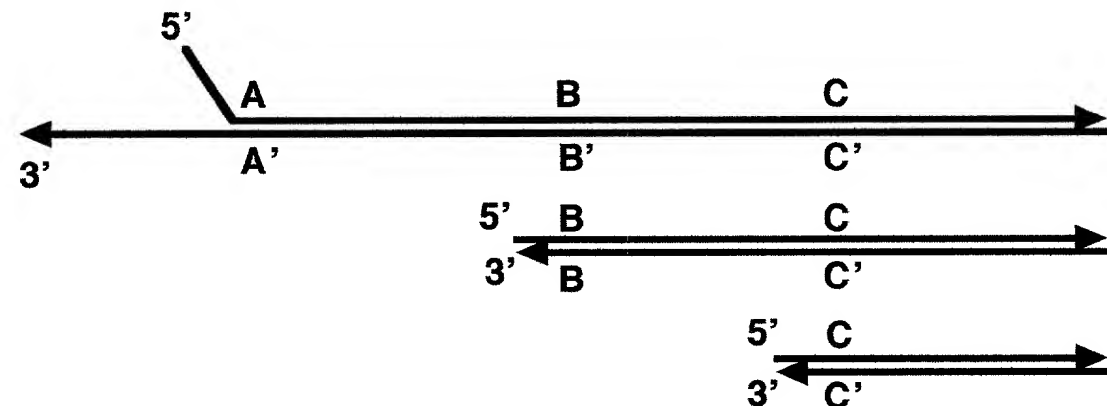
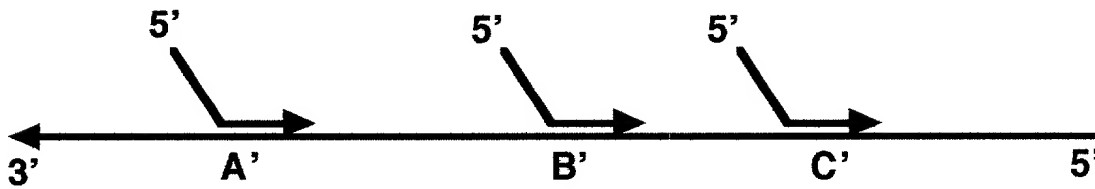


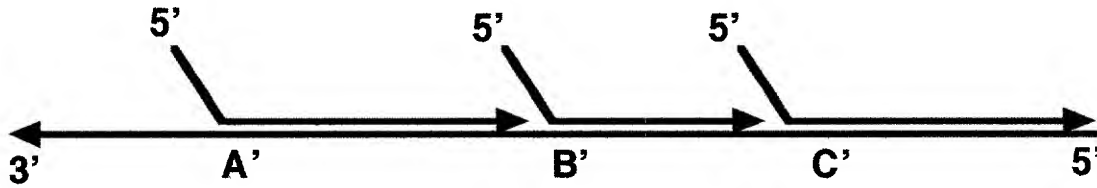
FIGURE 4

TOPPERS™ 26296860

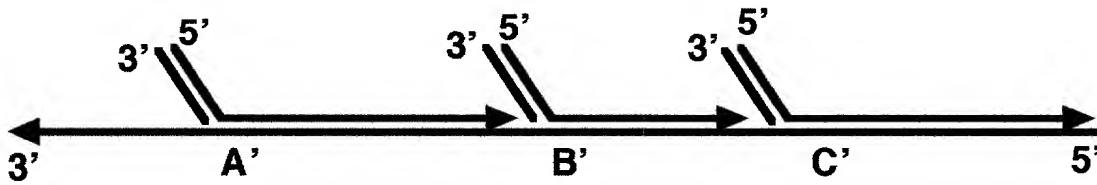
(1) Binding of random primers to 1st cDNA strand



(2) Extension of random primers using 1st cDNA strand as template



(3-a) Creation of functional promoters by binding of complementary strand



(3-a) Creation of functional promoters by self-complementary sequences

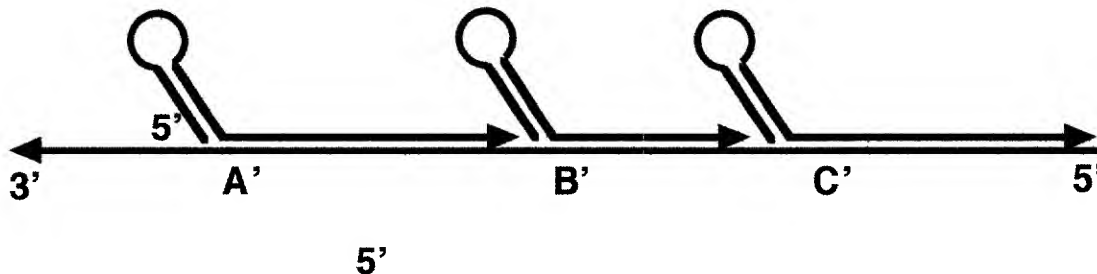
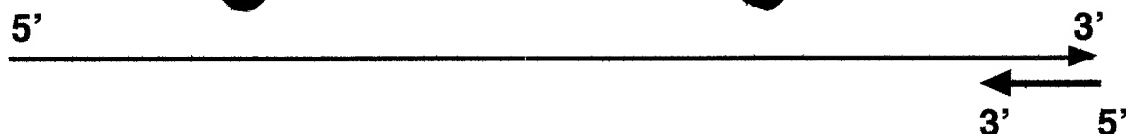
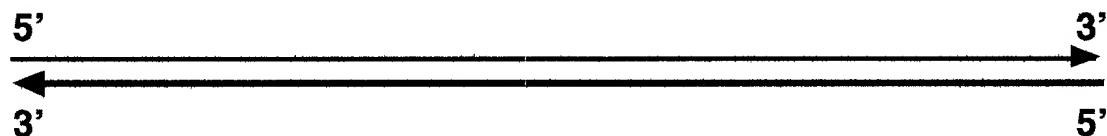


FIGURE 5

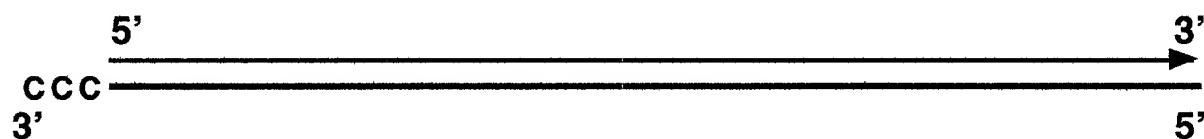
(A) Binding of Primer to analyte



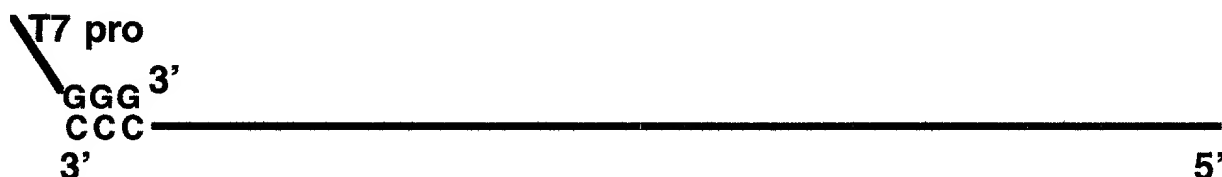
(B) Extension of Primer using analyte as template



(C) Template Independent addition of dCTP



(D) Use of 3' end of 1st cNA strand for binding of Primer with T7 promoter



(E) Binding of Primer with T7 promoter to internal sequenced of cNDNA

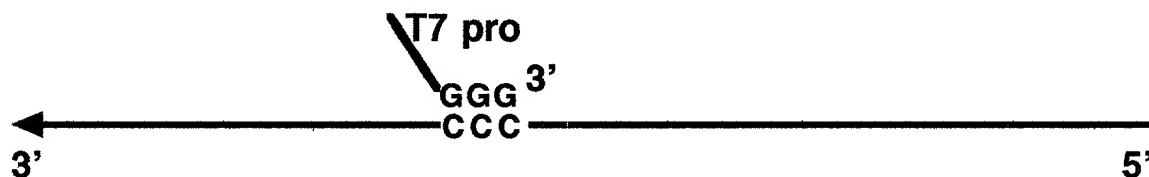


FIGURE 6

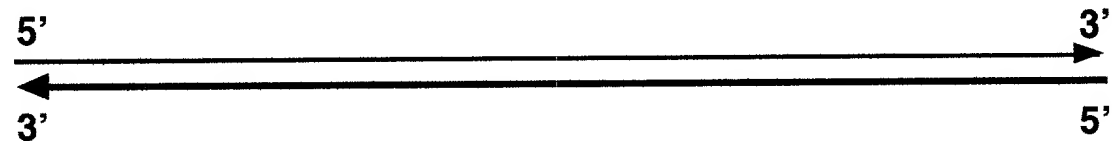
(A) RNA Substrate



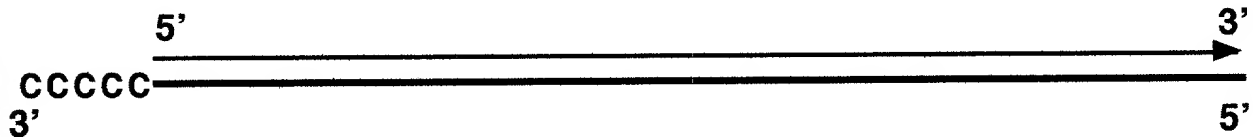
(B) Binding of Primer to RNA Substrate



(C) Extension of Primer using RNA as template



(D) Template Independent addition of dCTP by Terminal Transferase



(E) Use of 3' end of 1st cDNA strand for binding of Primer with T7 promoter

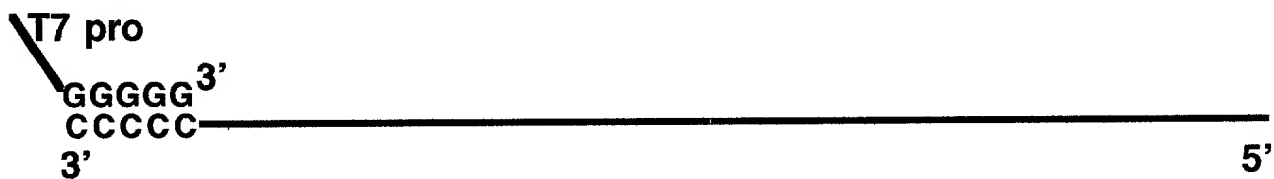
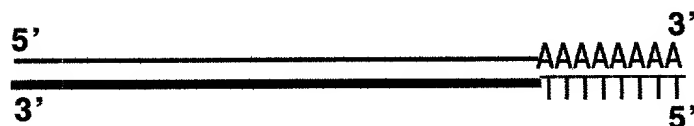


FIGURE 7

1) analyte



2) cNA copy made from analyte



3a) double-stranded oligonucleotide ligated to RNA/DNA hybrid by T4 DNA ligase



3b) single-stranded oligonucleotide ligated to a single-stranded 3' tail by T4 RNA ligase



3c) double-stranded oligonucleotide ligated to single-stranded 3' tail by T4 DNA ligase

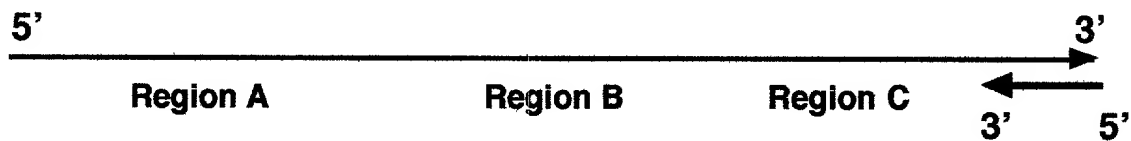


FIGURE 8

(A) RNA Substrate



(B) Binding of Primer to RNA Substrate



(C) Extension of Primer using RNA as template



(D) Nicking of cDNA strand followed by release from RNA template



(E) Template independent addition of dCTP and binding of primer with T7 Promoter

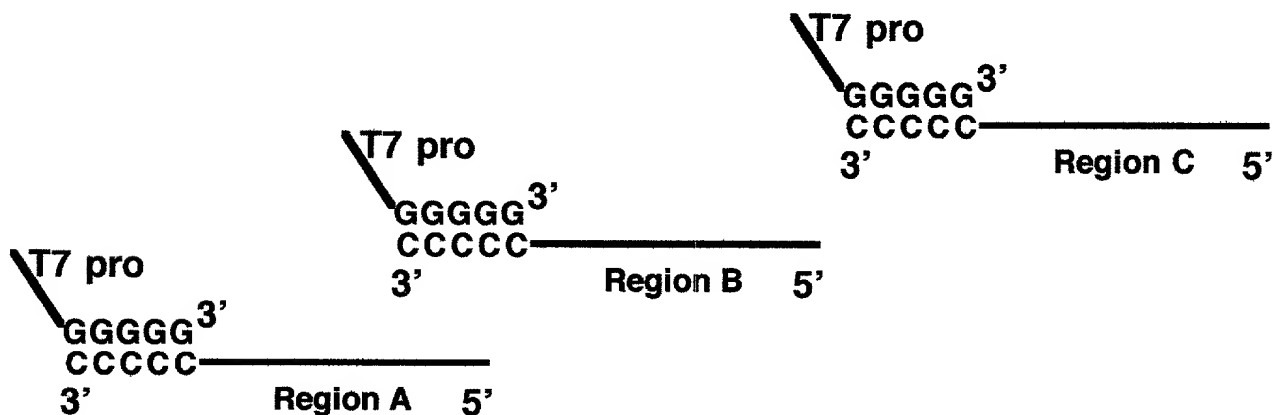


FIGURE 9

Downloaded from www.sciencedirect.com

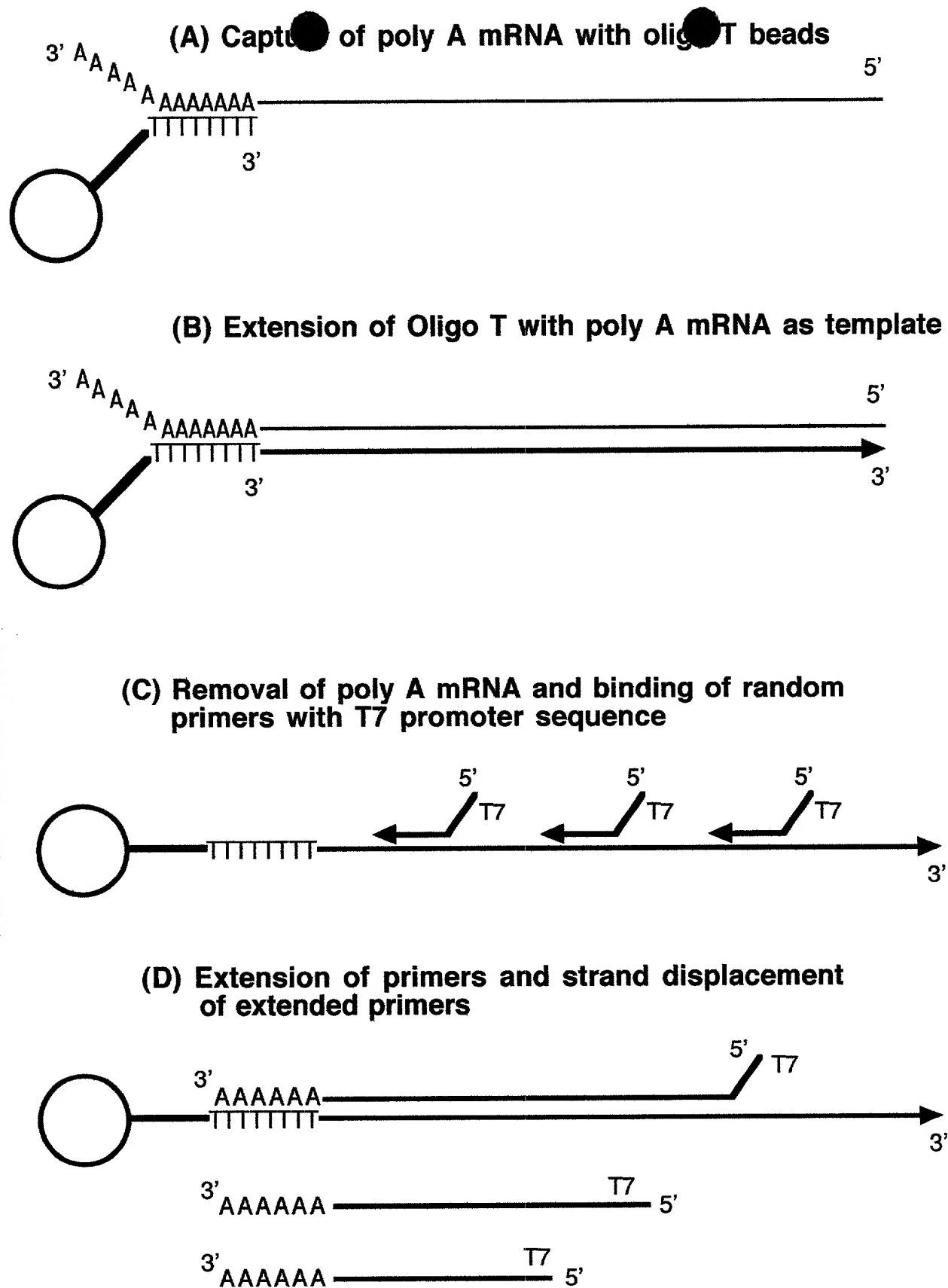
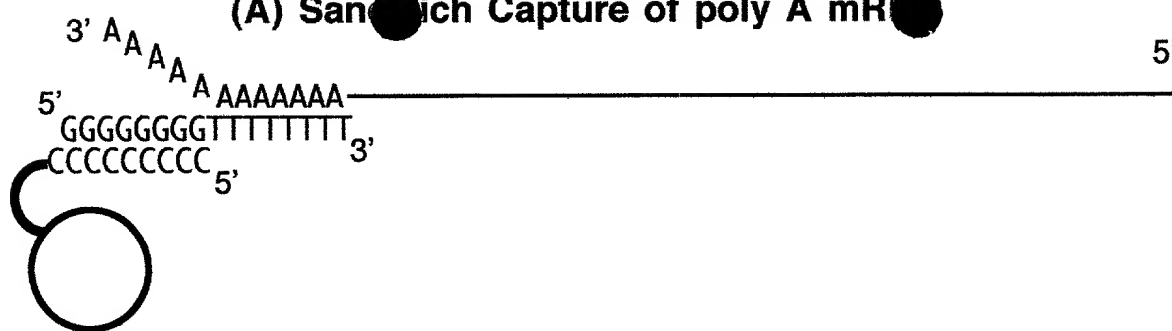
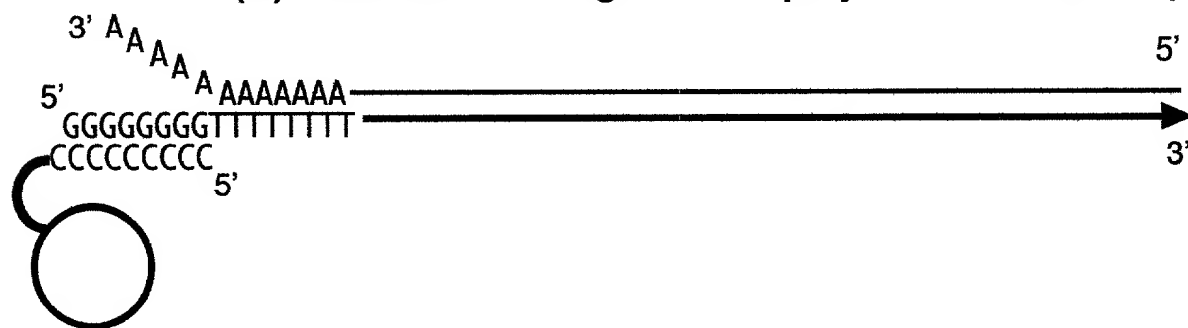


FIGURE 10

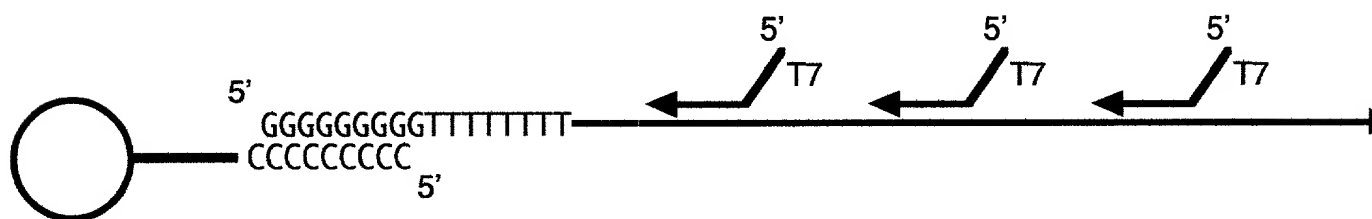
(A) Sandwich Capture of poly A mRNA



(B) Extension of Oligo T with poly A mRNA as template



(C) Removal of poly A mRNA and binding of random primers with T7 promoter sequence



(D) Extension of primers and strand displacement of extended primers

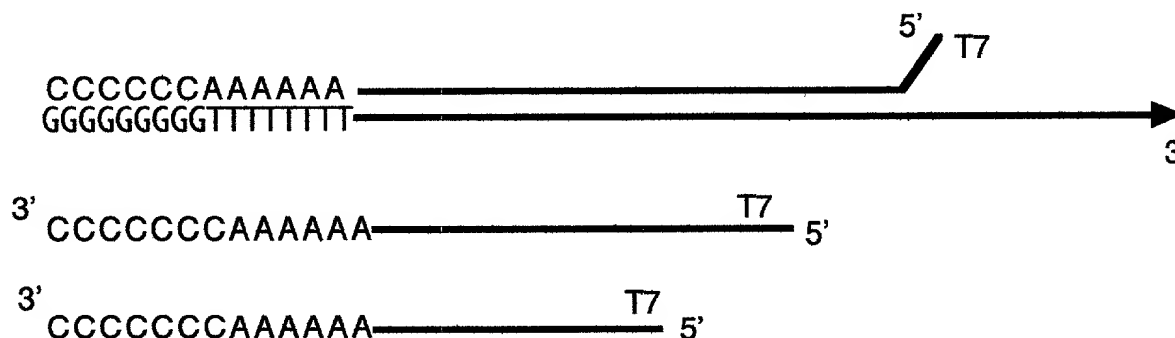


FIGURE 11

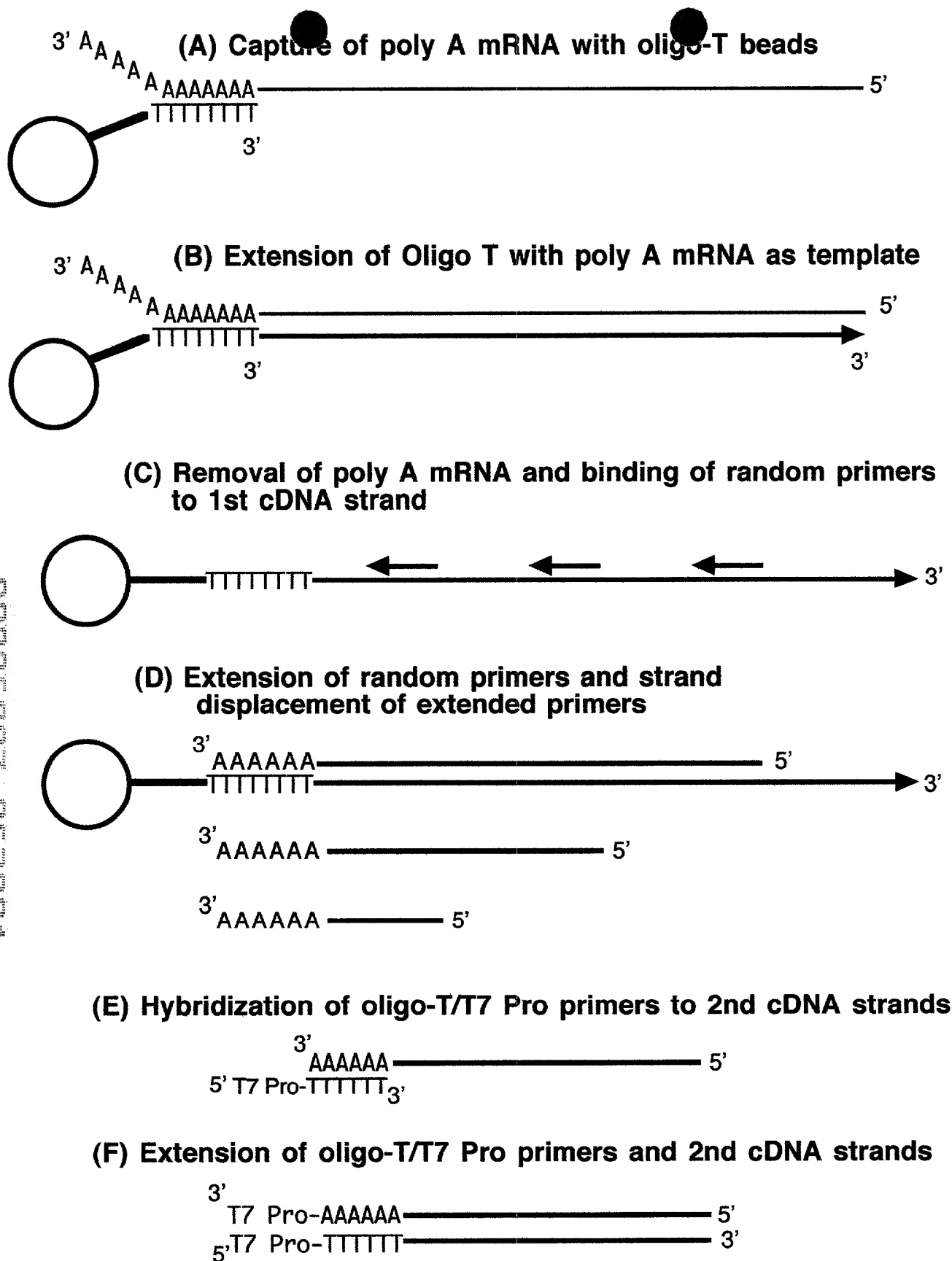
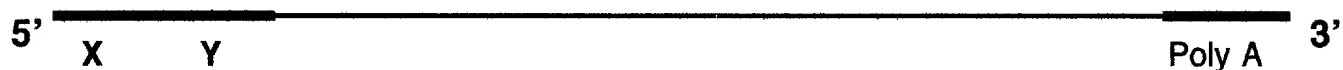


FIGURE 12

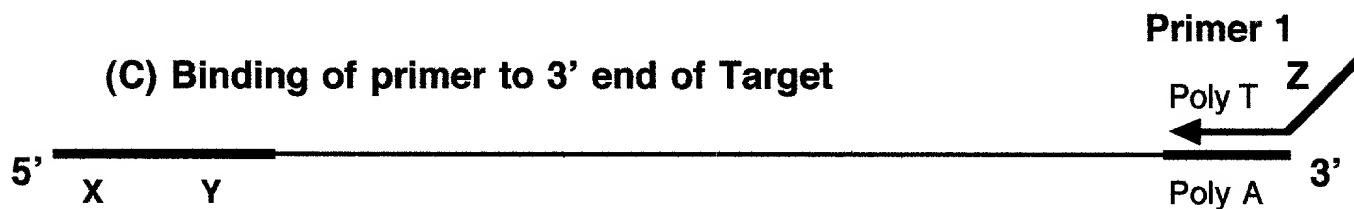
(A) Poly A RNA Target



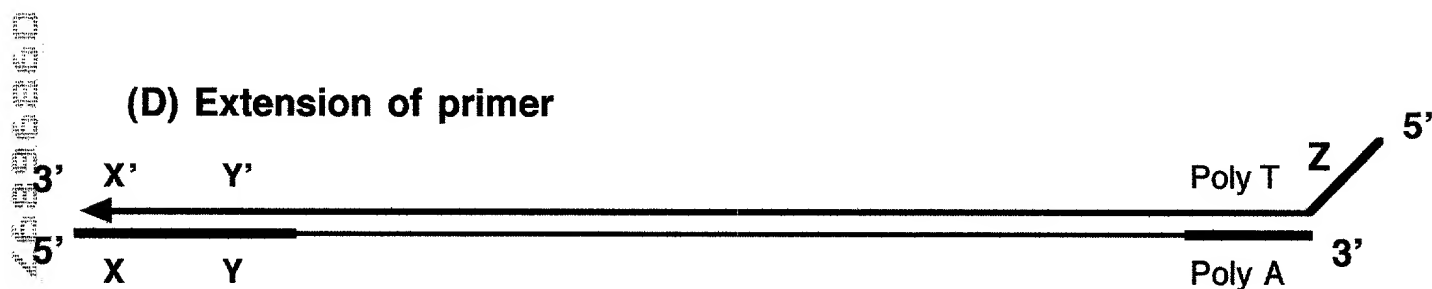
(B) Ligation of UDT to 5' end of Target



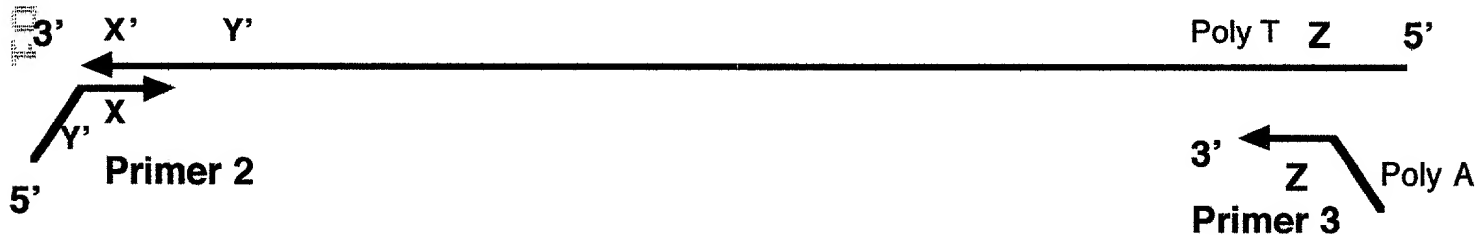
(C) Binding of primer to 3' end of Target



(D) Extension of primer



(E) Addition of Primers for Isothermal Amplification



(F) Unit length Isothermal Amplification

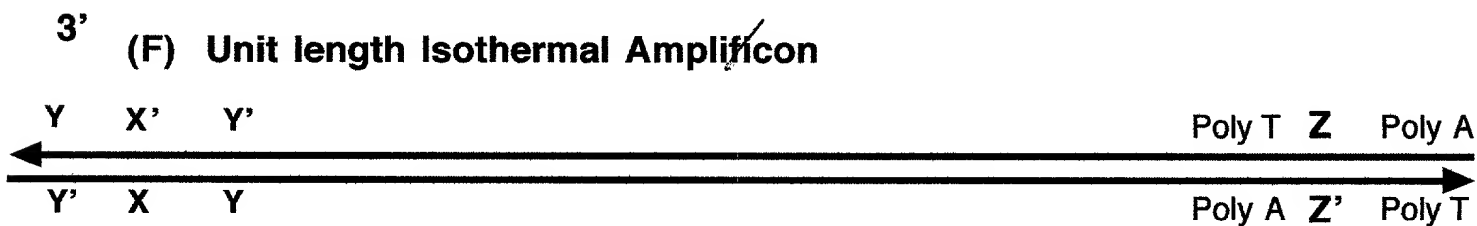


FIGURE 13

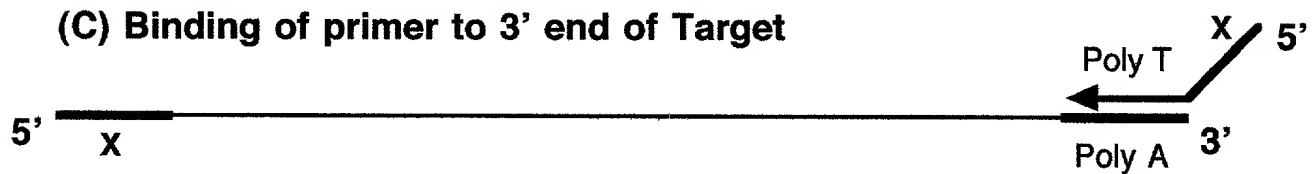
(A) Poly A RNA Target



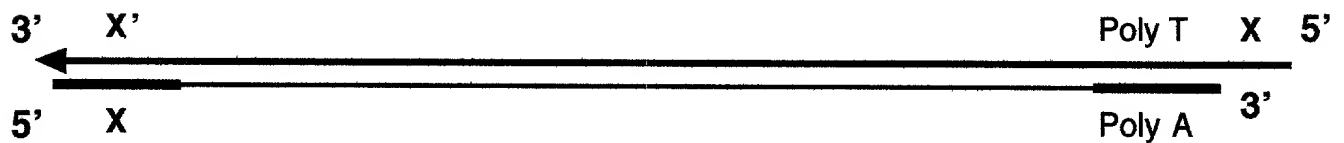
(B) Ligation of UDT to 5' end of Target



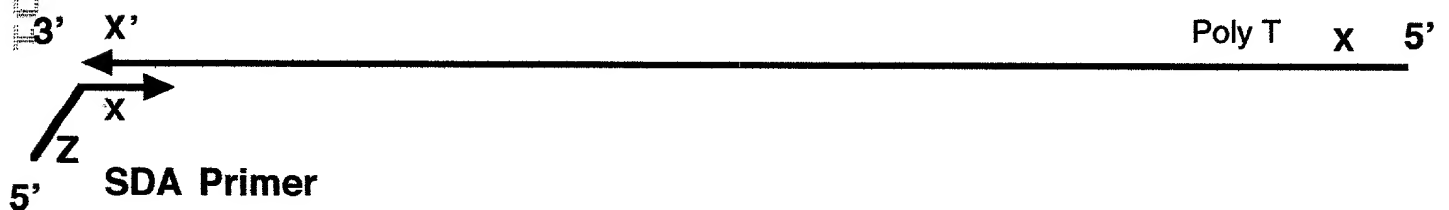
(C) Binding of primer to 3' end of Target



(D) Extension of primer



(E) Addition of SDA Primer



(F) Unit length SDA Amplificon

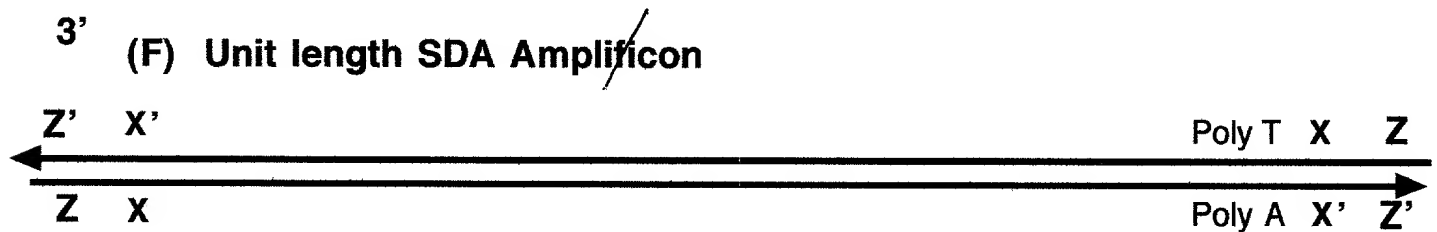
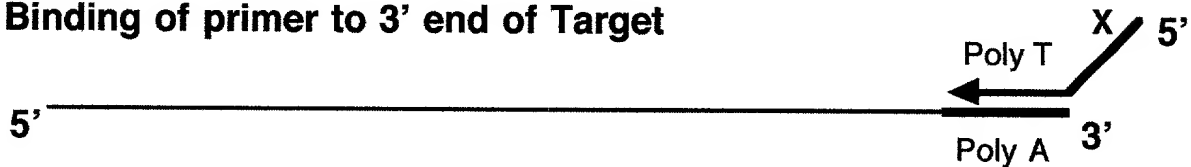


FIGURE 14

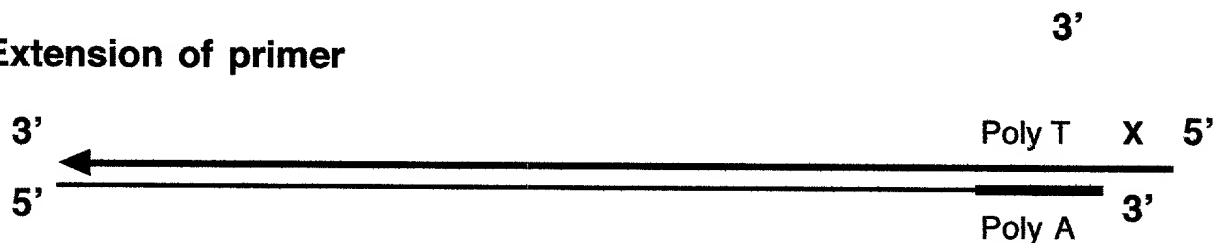
(A) Poly A RNA Target



(B) Binding of primer to 3' end of Target



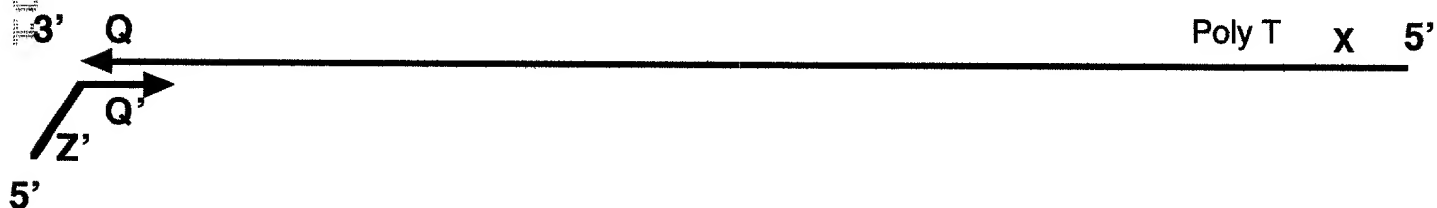
(C) Extension of primer



(D) addition of ^{UT}UTE (Q) to 3' end of first copy



(E) Addition of Primer for binding to Q



(F) Unit length Amplicon

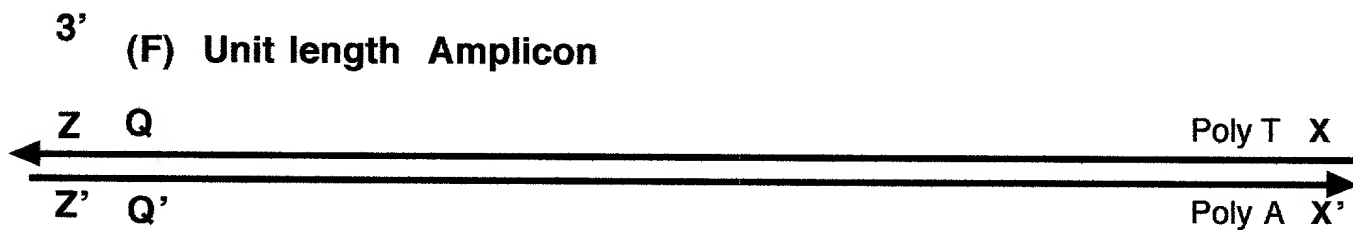
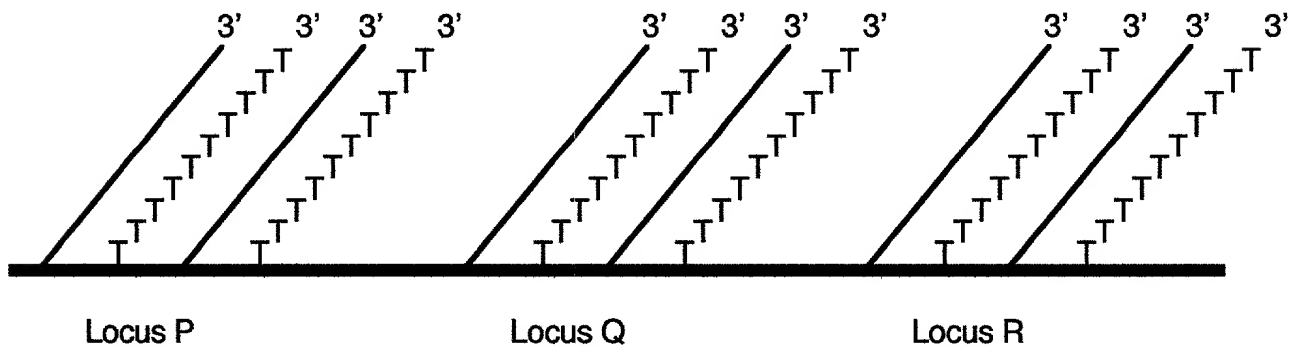


FIGURE 15

- (1) Array with SPE's complementary to analyte "P" at Locus P, SPE's complementary to analyte "Q" at Locus Q and SPE's complementary to analyte "R" at Locus R and with UPE's comprising Poly T sequences at all three loci



- (2) Binding of analyte "P" to corresponding SPE at Locus P

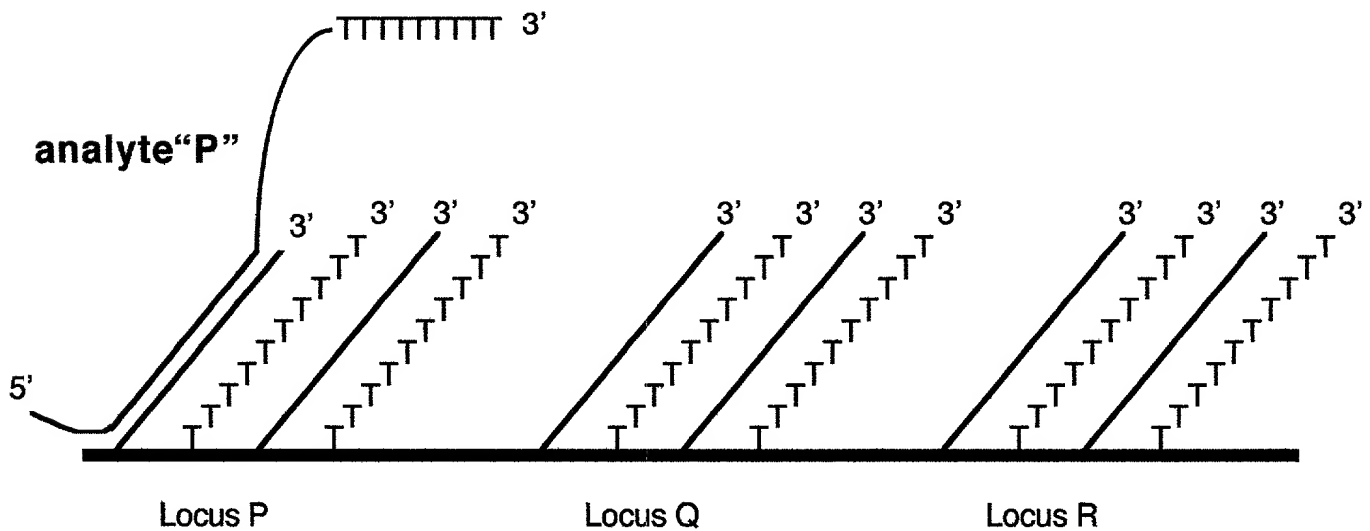


Figure 16

Binding of an analyte to an array with SPE's and UPE's

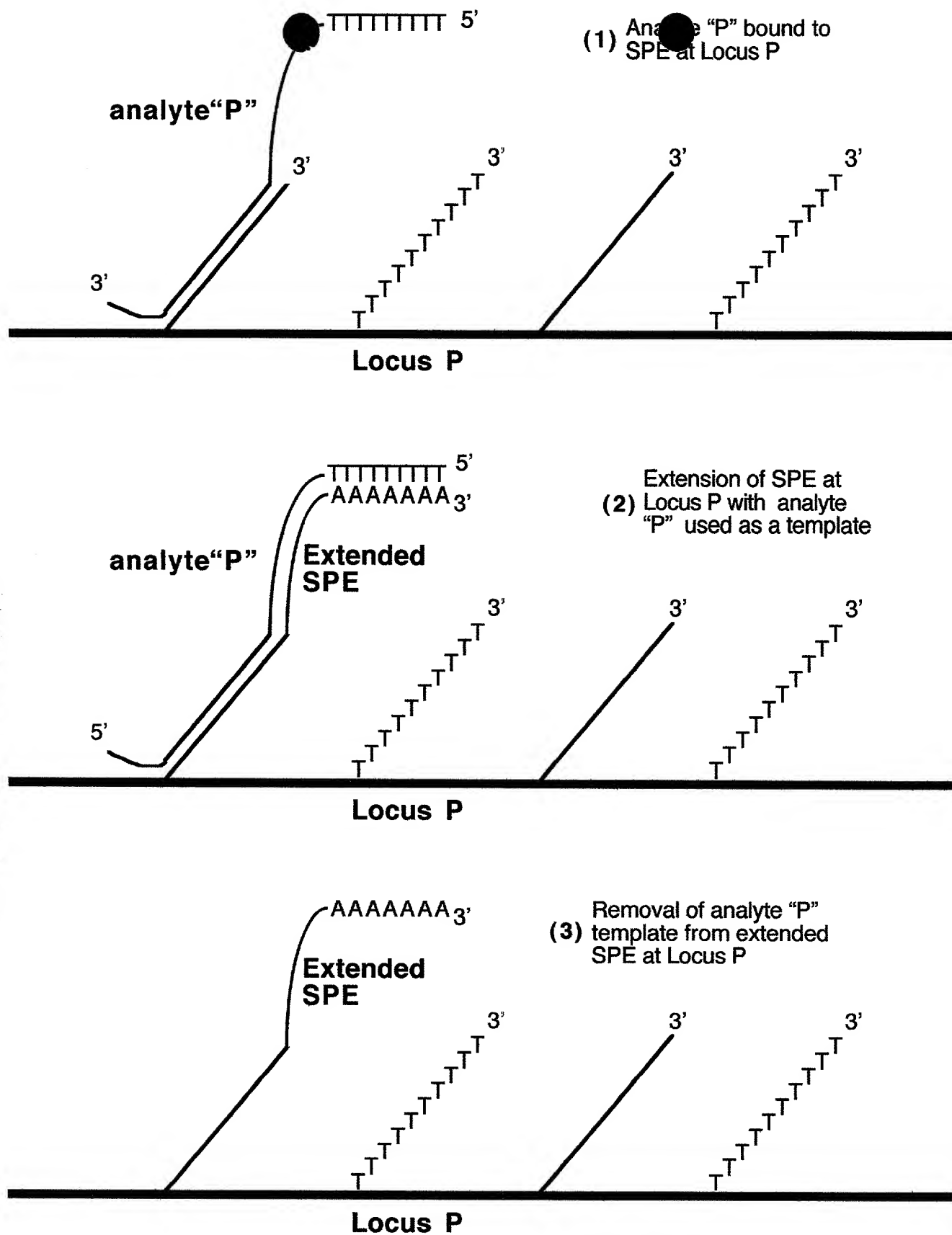


Figure 17
Extension of an SPE

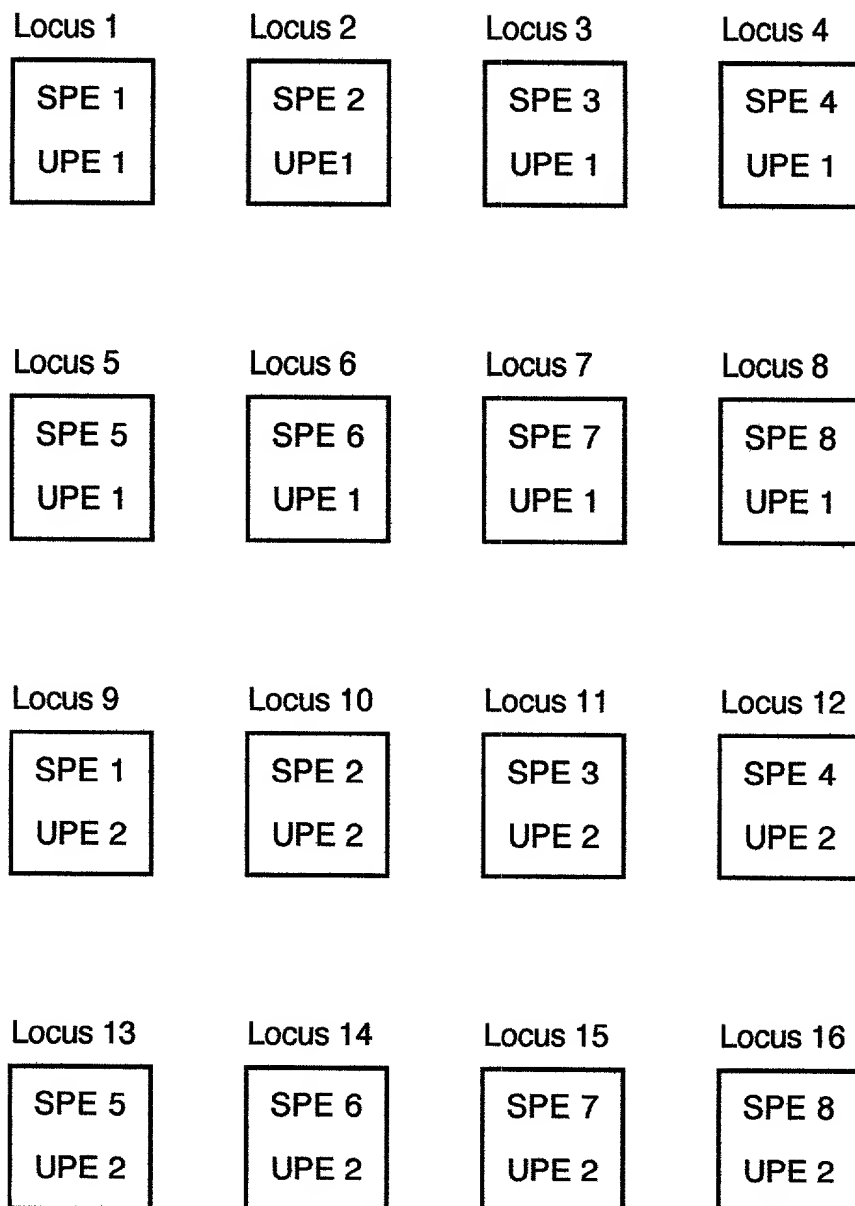
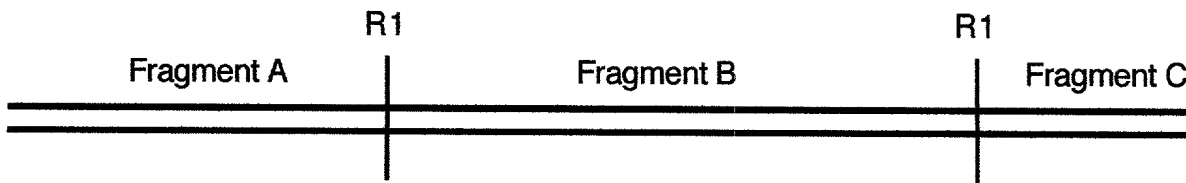


Figure 20
Amplification Array for Comparative Analysis

(1) Digestion of DNA with restriction enzyme R1



(2) Ligation of UDE's to DNA fragments



(3) Binding and extension of SPE primers with different 3' ends followed by extensions with UPE primers

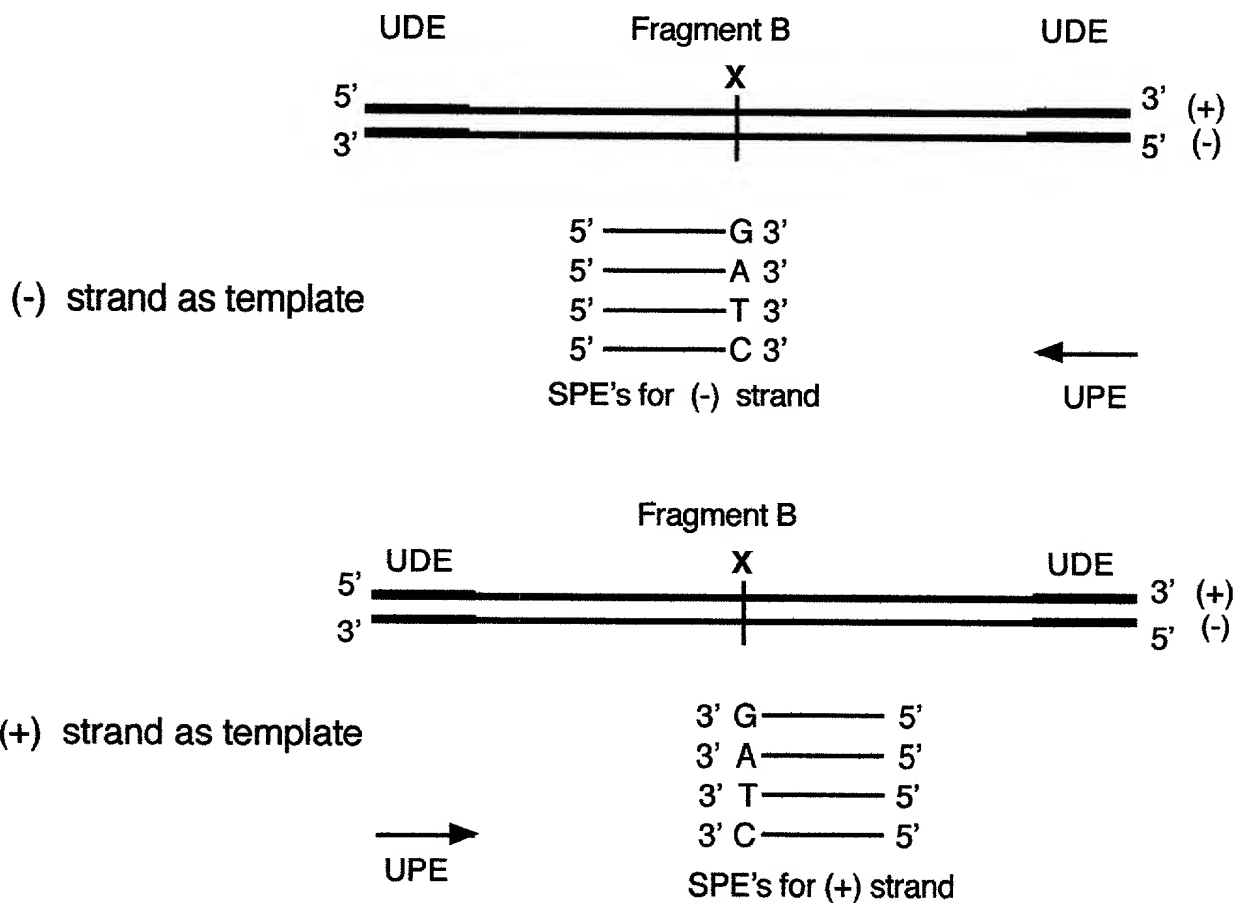


Figure 21

Use of an array with SPE's and UPE's for SNP analysis

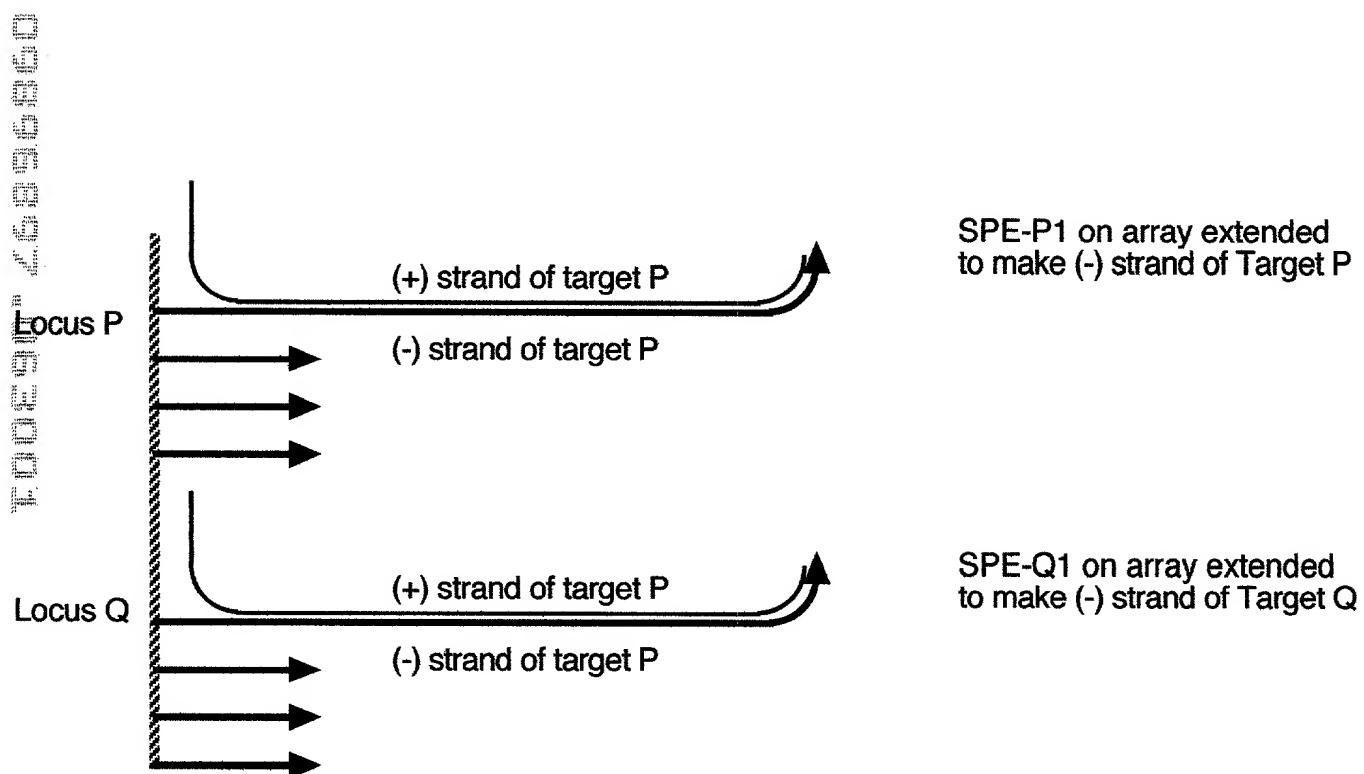
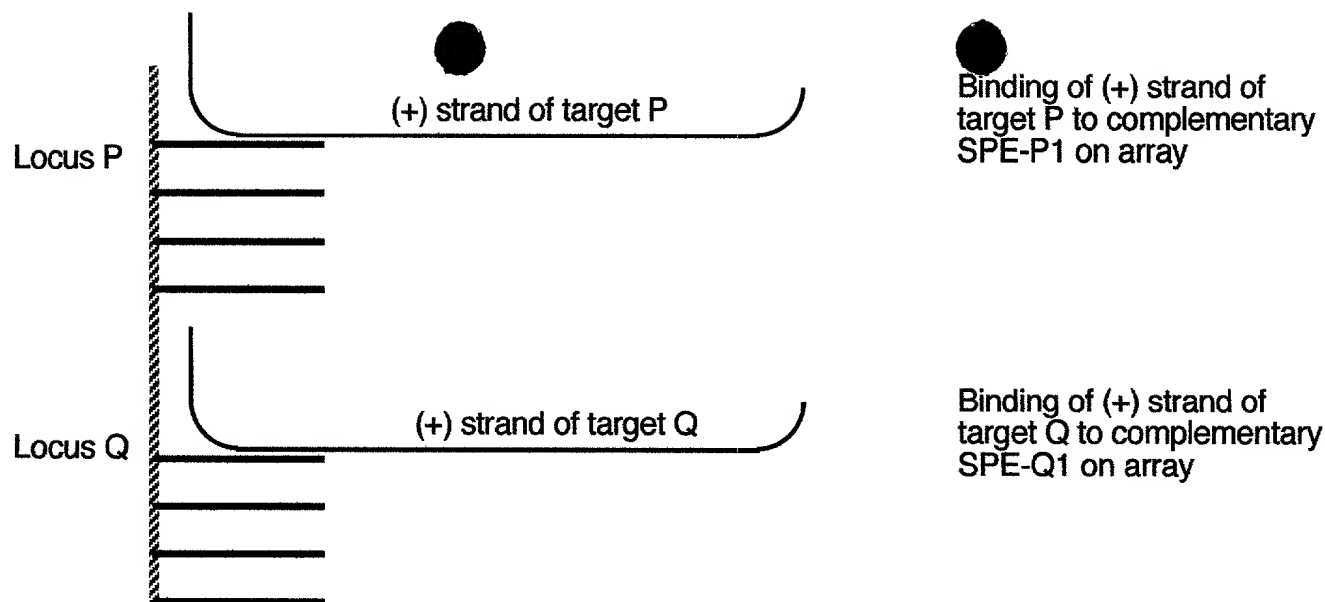


Figure 22

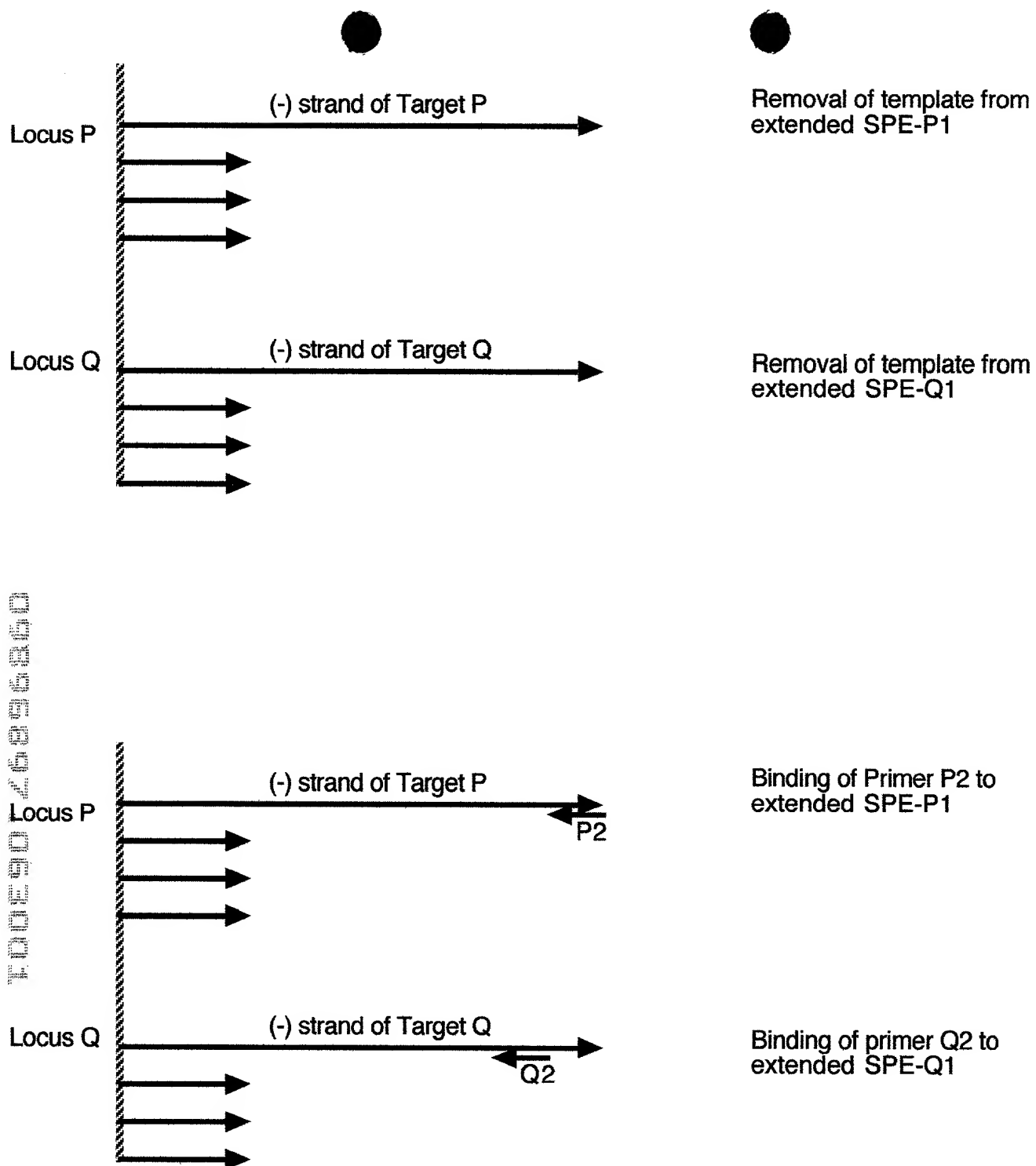
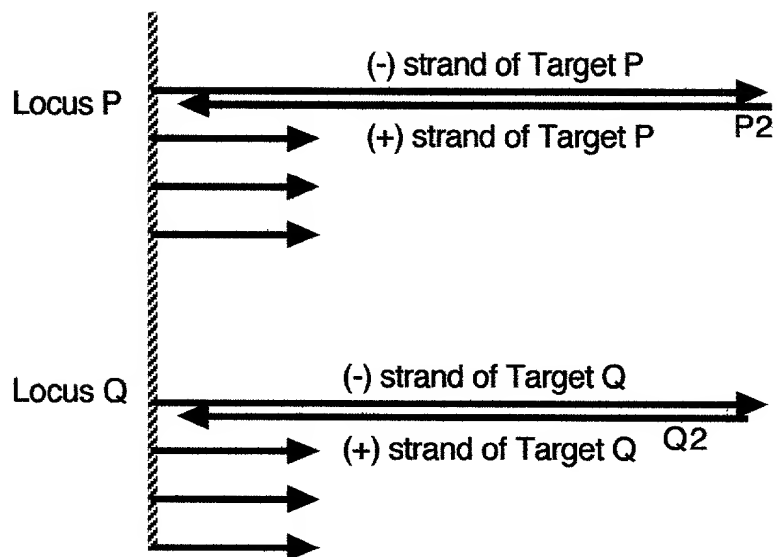
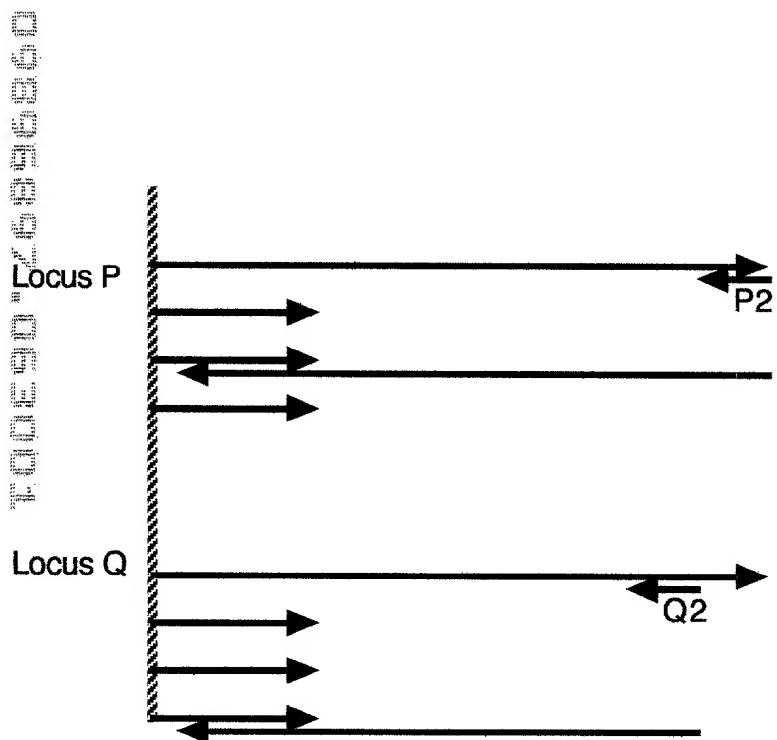


Figure 23



Extension of primer P2 by using extended SPE-P1 as a template

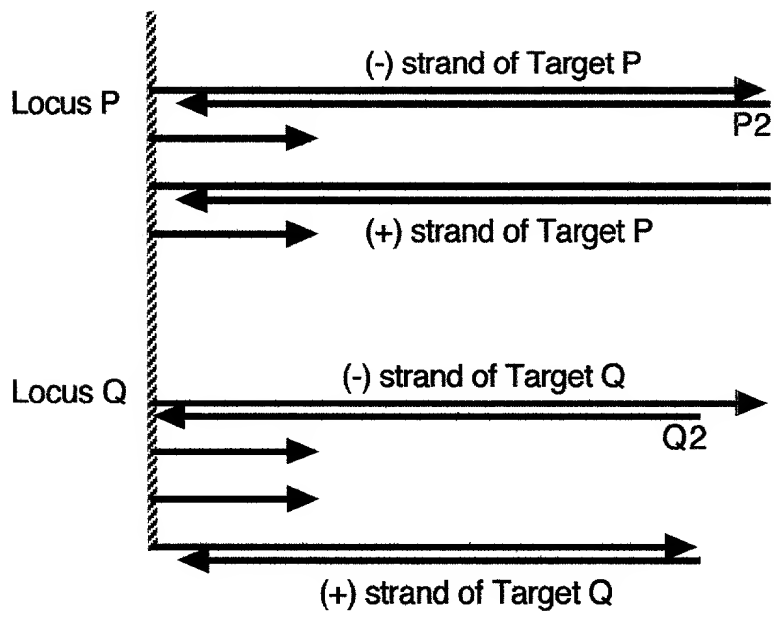
Extension of primer Q2 by using extended SPE-Q1 as a template



Denaturation followed by annealing of primer P2 to extended SPE-P1 and hybridization of extended P2 to un-extended SPE-P1

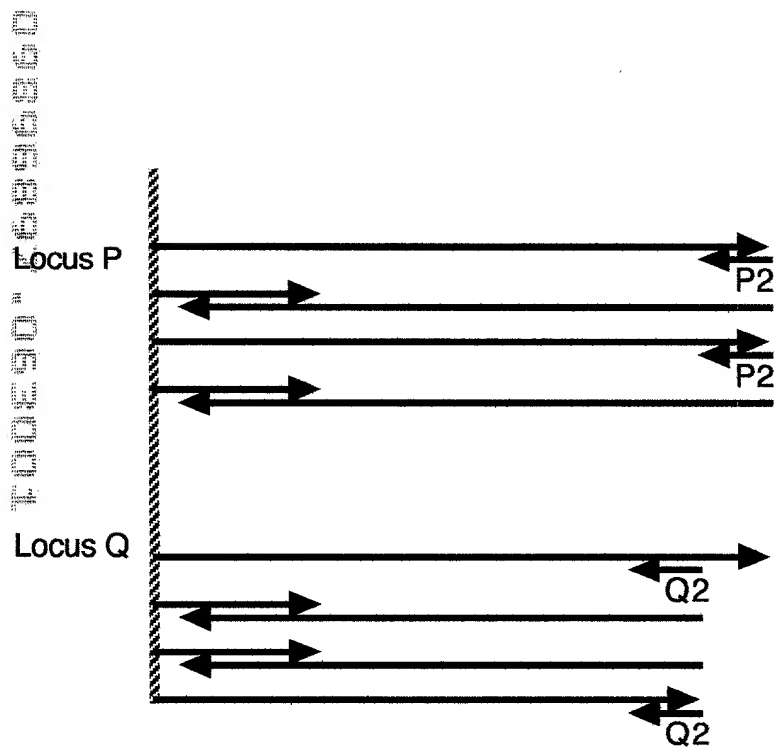
Denaturation followed by annealing of primer Q2 to extended SPE-Q1 and hybridization of extended Q2 to un-extended SPE-Q1

Figure 24



Extension of primer P2 and
SPE-P1

Extension of primer Q2 and
SPE-Q1



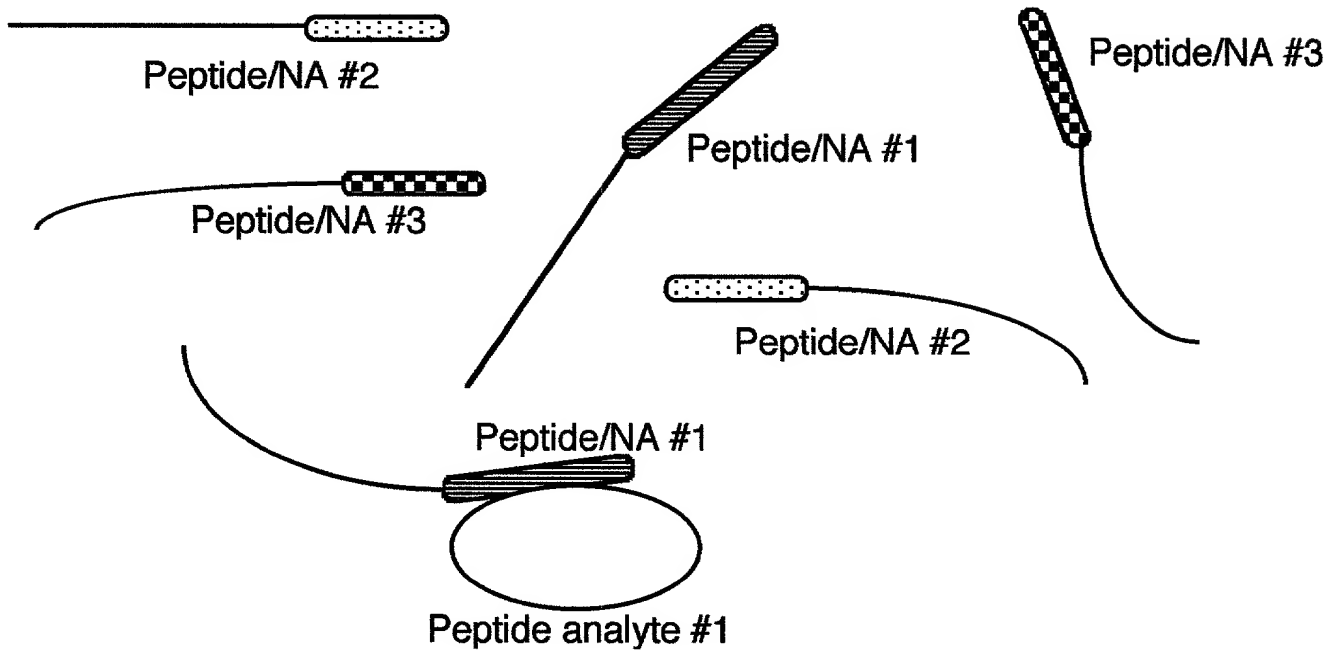
Denaturation followed by annealing of
primer P2's to extended SPE-P1's
and hybridization of extended P2's to
un-extended SPE-P1's

Denaturation followed by annealing of
primer Q2's to extended SPE-Q1's
and hybridization of extended Q2's to
un-extended SPE-Q1's

Figure 25

A) Mixture of a Library of Peptide analytes with a Library of Peptide/NAs

B) Binding of Peptide analyte #1 to Peptide/NA #1



C) Binding of Peptide/NAs to matrix through complementary sequences

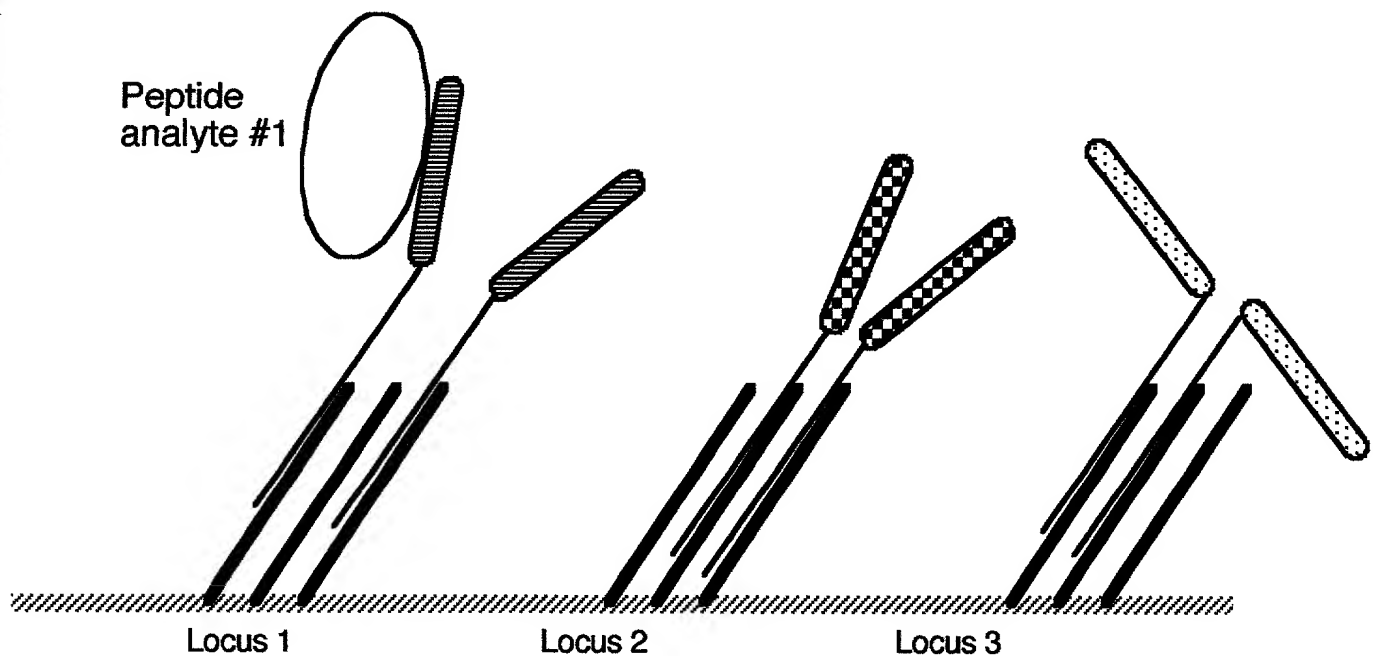


Figure 26

1 2 3 4 5

2000
1650
1000
850
650
500
400
300
200
100

- 2000
1650
1000
850
650
500
400
300
200
100

Figure 27

TOP-90-2639590

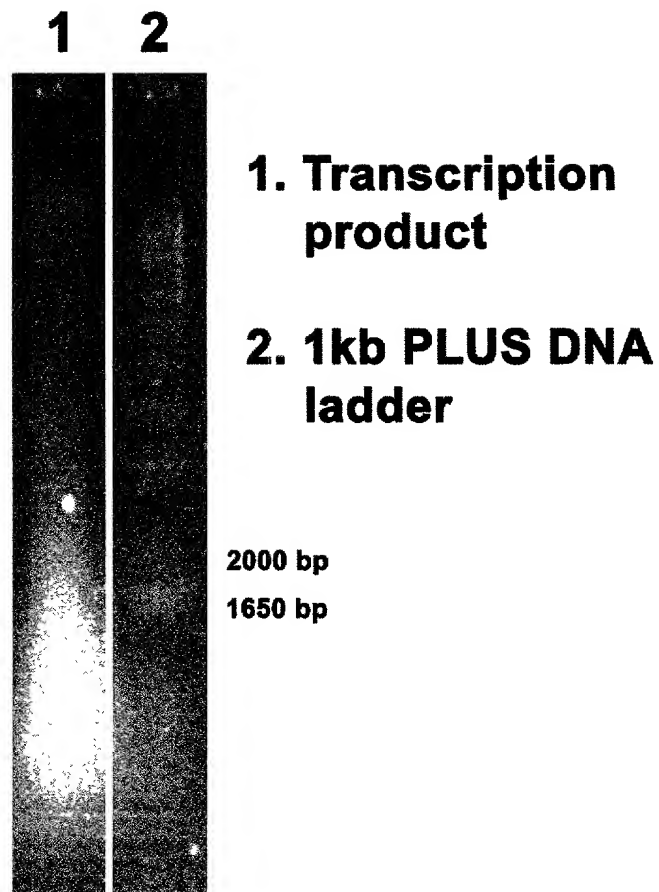


Figure 28

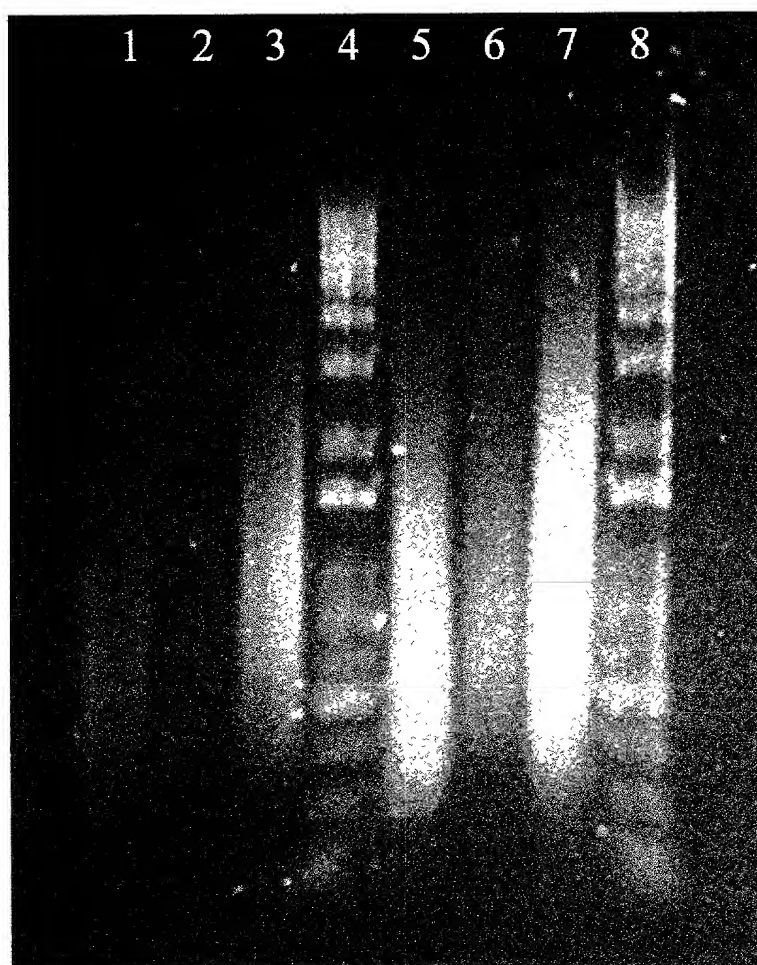
1 2



**1. Transcription
Product**

**2. 1 kb PLUS DNA
Ladder**

Figure 29



1. Random primers - 2 μ l

2. T7-C9 primers
without TdT tailing - 2 μ l

3. T7-C9 primers
after TdT tailing - 2 μ l

4. 1 kb PLUS DNA Ladder

5. Random primers - 10 μ l

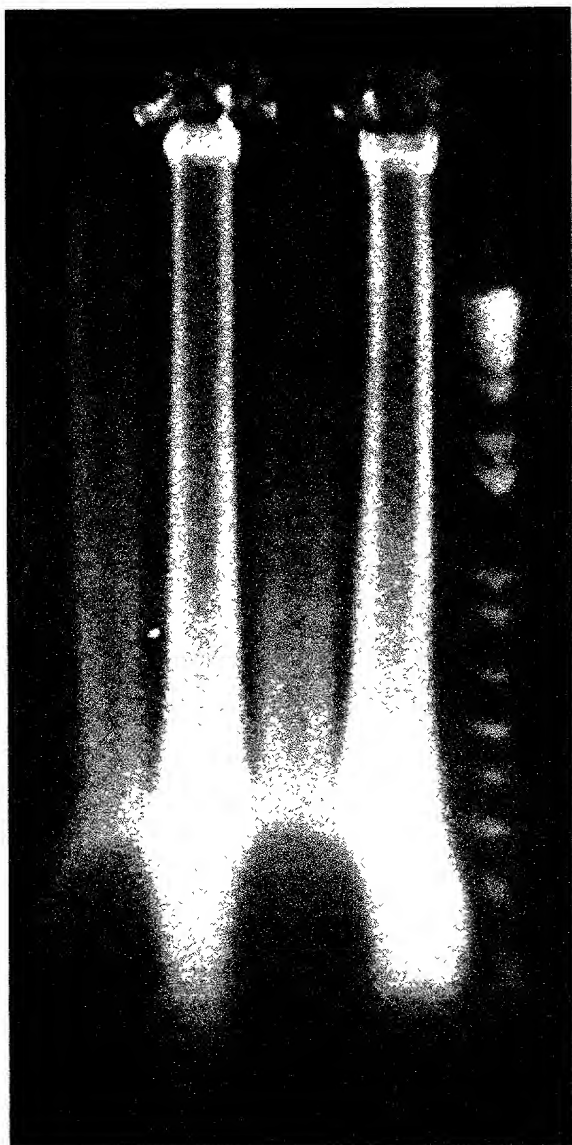
6. T7-C9 primers
without TdT tailing - 10 μ l

7. T7-C9 primers
after TdT tailing - 10 μ l

8. 1 kb PLUS DNA Ladder

Figure 30

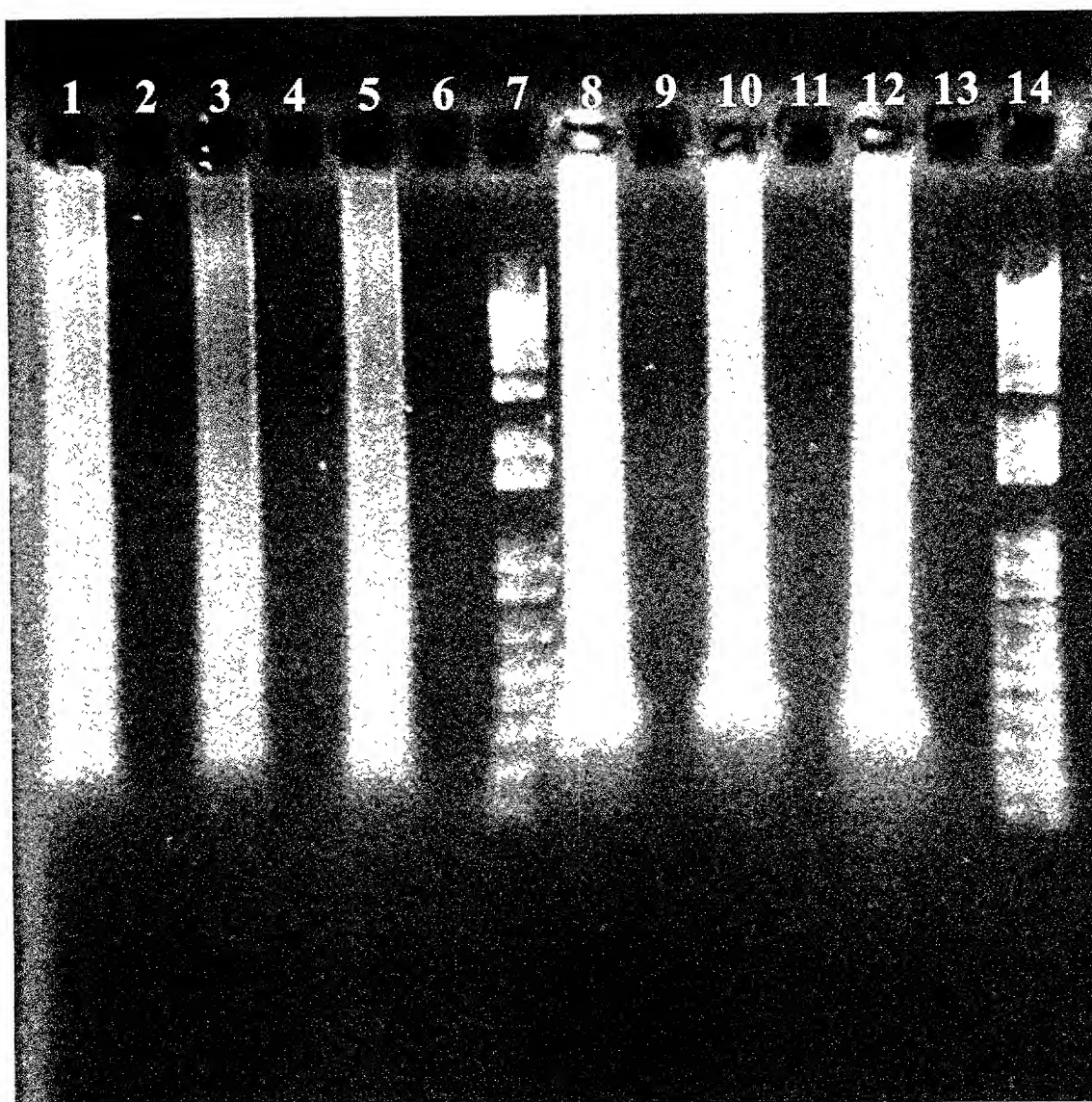
1 2 3 4 5



- 1. Taq pol. 1 cycle
- 2. Taq pol. 5 cycles
- 3. Tth pol. 1 cycle
- 4. Tth pol. 5 cycles
- 5. 1 kb PLUS DNA Ladder

Figure 31

46836350



1. Sample 1 - 4 μ l transcription product
2. Sample 1 - 1 μ l DNA template
3. Sample 2 - 4 μ l transcription product
4. Sample 2 - 1 μ l DNA template
5. Sample 3 - 4 μ l transcription product
6. Sample 3 - 1 μ l DNA template
7. 1 kb PLUS DNA Ladder

8. Sample 1 - 10 μ l transcription product
9. Sample 1 - 2.5 μ l DNA template
10. Sample 2 - 10 μ l transcription product
11. Sample 2 - 2.5 μ l DNA template
12. Sample 3 - 10 μ l transcription product
13. Sample 3 - 2.5 μ l DNA template
14. 1 kb PLUS DNA Ladder

Figure 32

1. 1 kb PLUS DNA Ladder
2. - - - -
3. Superscript II (Life Technologies)
4. M-MuLV (Life Technologies)
5. M-MuLV (New England Biolabs)
6. Enhanced AMV (Sigma)
7. AMV (Life Technologies)
8. AMV (Sigma)
9. Omniscript (Qiagen)
10. displayTHERMO-RT (Display systems Biotech)
11. Powescript (Clontech)
12. - - - -
13. λ - Hind III marker

Figure 33